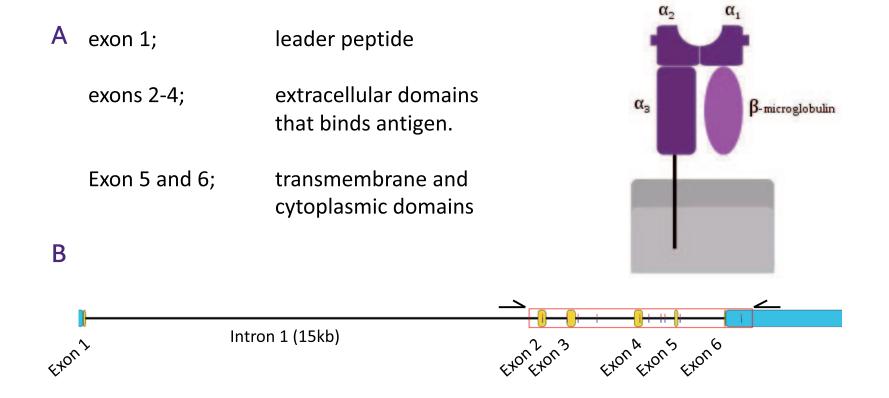
## GENDX

# The MR1 gene encompasses at least 6 alleles

Non-classical MHC class I genes, also known as MHC class Ib genes and MHC-I like genes, have few alleles and are considered oligo- or even mono-morphic. We developed a MR1 specific PCR assay and sequenced DNA samples from a diverse set of HLA genotypes. In this relatively small panel we found multiple alleles encoding for different MR1 proteins. Here we present new allele variants for MR1 and map the variant residues to the protein structure.

with MR1 presenting the drug diclofenac or its metabolite 5-OH-diclofenac 11. Accordingly, R9H might be relevant for TCR recognition of MR1 ligands whose presentation is not disrupted by R9H. E52G and H90Q are non-conservative changes and could be of consequence to the function or folding of MR1 but this should be tested in functional studies. I244V represents a conservative change in a region that is not known and unlikely to be of functional importance. R304K represents a conservative change in



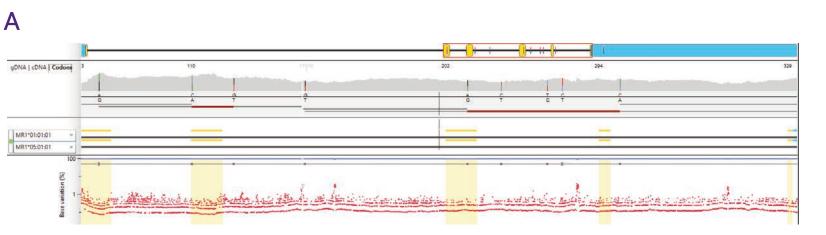
#### Materials & Methods

MR1 is a MHC-I like gene that is on chromosome 1. The structure of MR1 is that of a typical MHC class I gene, with exon 1 coding for a leader peptide and exons 2-4 coding for extracellular domains that can bind an antigen. Exon 5 and 6 are coding sequences for the transmembrane and cytoplasmic domains (Figure 1A). An unusual feature of MR1 is a large intron 1 of 15kb. To study variation of MR1 we developed a PCR assay that amplifies a region spanning intron 1 to the 3' UTR, so that the resulting amplicon of 8 kb includes all exons encoding for the MR1 protein (Figure 1B). We performed PCR reactions for 56 DNA samples from the genetic testing reference material from the Coriell Cell Repositories.

#### Sequencing results

We sequenced the 56 amplicons on an Illumina Miseq and analyzed the data using NGSengine software (Figure 2A). The sequence data was of high quality. A diverse set of heterozygous genotypes was discovered (Figure 2B). In all cases the minor fraction of the heterozygous positions was between 43-50%, indicating a balanced amplification of both alleles in each sample.

the cytoplasmic tail that has been observed in other species as well.

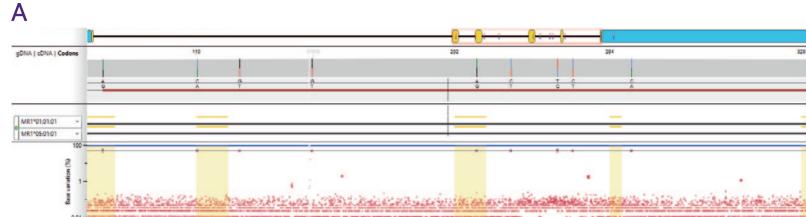


[Exon+] ESA% [Amplicon] HP 33107 NA17219 1.63 28440 NA17222 1,79 48,19 VA10005 22940 1,64 NA17226 29321 2,19 VA12244 19904 47.04 NA17228 30207 49,28 1,97 2,92 47,77 A16688 29860 NA17229 29556 1,12 A16689 26998 NA17230 26944 1,73 48.12 A17019 24333 45.96 NA17232 30134 A17020 23918 1,58 NA17234 30441 1,85 VA17039 29395 19567 1,88 46,75 3NA17240 A17057 27034 44,1 17483 1,92 A17058 27231 44,22 1,41 NA17246 17055 88 2,06 A17078 23598 2,63 43,79 1,59 NA17247 15286 48,45 46,28 A17084 21898 46,9 NA17256 20780 1,48 49.6 A17114 | 17885 48,33 NA17261 17234 2,06 47,54 NA17115 19316 NA17264 20280 1,59 48,77 47,48 A17119 | 18755 INA17267 18975 1,42 41,48 A17129 23455 45,55 NA17269 22885 A17130 19493 NA17274 24179 NA17201 20172 46,58 3 NA17282 20639 47,23 1,36 VA17203 19119 44 92 NA17285 23421 1,39 NA17204 26351 1,86 NA17286 25701 87 1,41 NA17205 25495 47,49 1,39 NA17289 23993 90 A17206 23477 NA17290 27217 NA17210 27078 NA17295 26253 1,61 1,09 90 NA17211 28286 2,48 NA17296 28872 1,37 NA17212 29607 3NA17440 25068 90 1,64 46.49 NA17213 27959 1 NA17466 23387 48,94 88 1,68 NA17214 34375 NA17618 23189

Figure 2: Considerable heterozygous positions in MR1 in 56 sample panel. (A) NGSengine coverage and base variation plot (B) sequence quality metrics from illumine sequence data. Shown are the mappability % (Map%), exon region maximum noise %, exon region estimated second allele fraction (ESA %), exon region heterozygous positions (HP) and heterozygous positions in the complete amplicon region ([amplicon] HP). 0 heterozygous positions is colored green and >0 heterozygous positions is colored yellow.

A. Polymorphism in the a1/a2- domains

**B.** Polymorphism in the a3domain/cyptoplasmictail Figure 1: MHC class I-related protein 1 (MR1) shares great homology with MHC-I genes. (A) Protein structure (B) genomic structure and primer binding sites.



В MR1\*01 RTHSLRYFRLGVS MR1\*02 RTHSLRYFRLGVSDPIRGVPEFISVGYVDSHPITTYDSVTROKEPRAPWMAENLAPDHWERYTOLLRGW OOMFKVELKRLORHYNHSGSHTYORMIGCELLEDGSTTGFLOYAYDGODFLIFNKDTLSWLAVDN MR1\*03 RTHSI RYFRI GVSDPIHGVPFFISVGYVDSHPITTYDSVTROKEPRAPWMAENI APDHWERYTOLI RGW OOMEKVELKRI ORHYNHSGSHTYORMIGCELLEDGSTTGELOYAYDGOD MR1\*04 RTHSLRYF<mark>H</mark>LGVSDPI**R**GVPEFISVGYVDSHPITTYDSVTRQKEPRAPWMAENLAPDHWERYTOLLRGW QQMFKVELKRLQRHYNHSGSHTYQRMIGCELLEDGSTTGFLQYAYDGODFLIFNKDTLSWLAVDNV

MR1\*05 RTHSLRYFRLGVSDPIHGVPEFISVGYVDSHPITTYDSVTRQKEPRAPWMA<mark>G</mark>NLAPDHWERYTQLLRGWQQMFKVELKRLQRHYNHSGS<mark>Q</mark>TYQRMIGCELLEDGSTTGFLQYAYDGQDFLIFNKDTLSWLAVDNV. MR1\*06 RTHSLRYFRLGVSDPIHGVPEFISVGYVDSHPITTYDSVTRQKEPRAPWMAENLAPDHWERYTQLLRGW QQMFKVELKRLQRHYNHSGSHTYQRMIGCELLEDGSTTGFLQYAYDGQDFLIFNKDTLSWLAVDNVA

MR1\*01 HTIKQAWEANQHELLYQKNWLEEECIAWLKRFLEYGKDTLQRTEPPLVRVNRKETFPGVTALFCKAHG F MR1\*02 HTIKQAWEANQHELLYQKNWLEEECIAWLKRFLEYGKDTLQRTEPPLVRVNRKETFPGVTALFCKAHG FYPPEIYMTWMKNGEEIVQEIDYGDILPSGDGTYQAWASIELDPQSSNLYSCH MR1\*03 HTIKOAWEANQHELLYQKNWLEEECIAWLKRFLEYGKDTLQRTEPPLVRVNRKETFPGVTALFCKAHG FYPPFIYMTWMKNGEEIVOEIDYGDILPSGDGTYQAWASIELDPOSSNLYSCH /R1\*05 HTIKQAWEANQHELLYQKNWLEEECIAWLKRFLEYGKDTLQRTEPPLVRVNRKETFPGVTALFCKAHG FYPPFIYMTWMKNGFFIVOFIDYGDII PSGDGTYOAWAS<mark>W</mark>FI DPOSSNI YSCHVFHCGVHMVI OVPI

VR1\*01 ESETIPLVMKAVSGSIVLVIVLAGVGVLVWRRRPREONGAIYLPTF MR1\*02 ESETIPLVMKAVSGSIVLVIVLAGVGVLVWRRRPREQNGAIYLPTPL MR1\*03 ESETIPLVMKAVSGSIVLVIVLAGVGVLVWRRRPREQNGAIYLPTP MR1\*04 ESETIPLVMKAVSGSIVLVIVLAGVGVLVWRRRPREQNGAIYLPTPI /R1\*05 ESETIPLVMKAVSGSIVLVIVLAGVGVLVWRRRPREQNGAIYLPTPD MR1\*06 ESETIPLVMKAVSGSIVLVIVLAGVGVLVWRR<mark>K</mark>PREQNGAIYLPTPDR

Figure 3: Six fully phased MR1 alleles. (A) NGSengine coverage and base variation plot from Pacbio sequence data from samples (B) novel MR1 protein variants.

To obtain fully phased MR1 allele sequences, MR1 was re-amplified from 5 samples containing alleles encoding for novel MR1 proteins. These amplicons were sequenced on a PacBio Sequel II System. This method resulted in highquality long reads that covered the complete amplicons and in which phasing of all heterozygous positions could be achieved (Figure 3).

#### Structure analysis

Figure 4 shows a cartoon display of MR1 with the variant residues in 5 novel alleles highlighted in teal. R9 represents a TCR contact in crystal structures of a MAIT TCR complexed

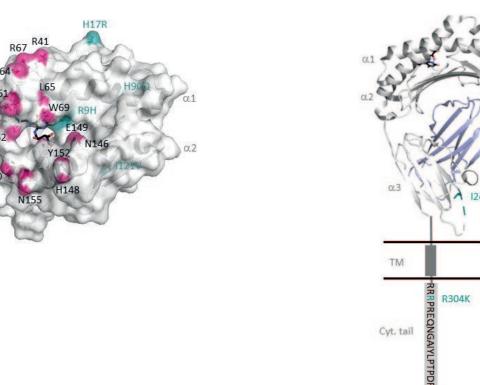


Figure 4: Cartoon and surface display of MR1 protein with variant positions

### Conclusion

Despite the monomorphic classification of MR1, a diverse set of heterozygous genotypes was discovered in a panel of 56 samples, including 5 new alleles encoding for protein variants. The data presented here is consistent with marked variation in MR1.

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